

Sarcopenia: An emphasis on occlusion training and dietary protein

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Abstract

Demographics reveal that the world's population aged 60 years and older will triple from 600 million in 2000 to more than 2 billion by the year 2050. To remain independent and healthy, an important factor to consider is the maintenance of skeletal muscle, as the elderly seem to become prone to a progressive loss of skeletal muscle with aging, termed sarcopenia. Interventions should focus on resistance training and optimal nutrition. Low intensity occlusion training may provide a mode of resistance training more applicable to the elderly, due to the lower loads used. Furthermore, an emphasis must be placed on high quality protein adequately distributed throughout the day to maximize protein synthesis. The use of drug therapy may be of some benefit, but it appears exercise and diet likely plays a more prominent role in the preservation of muscle mass and strength than administration of synthetic hormones. Hippokratia 2011; 15 (2): 132-137

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Demographics reveal that the world's population aged 60 years and older will triple from 600 million in 2000 to more than 2 billion by the year 2050. Due to an increased life expectancy, good health is essential for those aging, so they can remain independent and prevent or delay the onset of noncommunicable and chronic metabolic diseases such as heart disease, cancer, stroke and diabetes¹. To remain independent and healthy, an important factor to consider is the maintenance of skeletal muscle, as the elderly become prone to a progressive loss of skeletal muscle with aging, termed sarcopenia (defined as appendicular skeletal muscle mass / height² <7.26 kg/m for males and <5.45 kg/m for females)². This is of importance because skeletal muscle functions as the largest disposal site of ingested glucose³, plays an important role in lipid oxidation^{4,5}, and is one of the single greatest contributors to the resting metabolic rate (RMR)⁶.

Age related sarcopenia begins in the 4th decade of life and has numerous adverse consequences⁷⁻¹¹. The loss of muscle mass is associated with an increased risk of falls^{12,13} and the risk for all-cause mortality is inversely associated with strength levels¹⁴. The exact cause is multifactorial, however alpha-motoneurons appear largely responsible for the loss of muscle mass with aging¹⁵⁻¹⁷. In addition some evidence supports an age related desensitization to anabolic stimuli from exercise^{18,19} and nutrition interventions²⁰⁻²², compounded by a decline in overall dietary intake²³.

Muscle hypertrophy is thought to occur from an increased rate of protein synthesis that chronically exceeds the rate of protein breakdown. Due to the overall desensitization to anabolic stimuli, some research has focused

on the ubiquitin proteasome systems role with increasing protein breakdown. However, a thorough review details that much of this research has been completed in rodents and the conditions in which the studies were completed might not necessarily reflect the physiology of older humans²⁴. To illustrate, the ubiquitin ligases MAFbx/atrogen-1 and MuRF1 are activated in conditions of muscle wasting caused by inflammation (e.g. cancer, COPD, severe trauma) but the data is inconsistent in medically stable individuals²⁴. Furthermore, a previous study found that insulin-mediated alterations in the protein expression of atrogen-1 and MuRF1 resulted in no changes in muscle protein breakdown²⁵. This suggests that if the ubiquitin proteasome system is playing a role in sarcopenia, it is minimal.

As previously stated alpha-motor neuron loss appears largely responsible for the in the loss of muscle mass. The mechanisms behind this loss are not fully understood but could potentially be an age-related decline in the synthesis of a protein (ciliary neurotrophic factor) that promotes the differentiation and survival of alpha-motoneurons in a rodent model²⁶. In addition, alterations to the neuromuscular junction have also been proposed. Delbono²⁷ and Narici and Mafulli²⁸ have described in detail the degenerative changes, which include fragmentation in the distribution of acetylcholine receptors, and an increase in the incidence of branches or boutons that are spatially separate or only connected by fine nerve filaments, suggesting fragmentation of the terminal. Interestingly, there is evidence to suggest that alterations in the muscle cell can occur independently of those in the neuron. To illustrate, apoptosis of skeletal muscle may be linked to mito-

chondrial dysfunction which is thought to increase with advancing age²⁹. Mitochondrial dysfunction coupled with altered respiratory chain function and decreased cell defense against free radicals can cause mitochondrial DNA damage which has been associated with the development of sarcopenia³⁰.

The purpose of this review is to provide a unique look at a novel exercise modality, specifically for those patients who are contraindicated to high intensity exercise, and a dietary protein nutritional intervention which could help prevent or attenuate this age-related disease. Drug and hormonal therapy will also be briefly discussed as possible supplements to exercise and nutrition.

Low-Load Resistance Exercise Intervention

Resistance exercise provides a means of preserving skeletal muscle tissue with age. The positive effects of high intensity resistance training in the elderly population have been well documented³¹⁻³⁴. Despite the positive effects seen with high intensity exercise, many elderly are contraindicated to high load resistance training and are limited to low-load, low-intensity exercise³⁵; resulting in a failure to recruit fast twitch fibers leading to the 20-50% reduction in fast twitch cross sectional area (CSA) reported in the literature^{17,36}.

A novel mode of exercise has recently received a lot of attention in the scientific literature as it appears to contradict the resistance training position stand of the American College of Sports Medicine (ACSM). The American College of Sports Medicine (ACSM) recommends lifting a weight of at least 70% of one's one repetition maximum (1RM) to achieve muscular hypertrophy as it is believed that anything below this intensity rarely produces substantial muscle hypertrophy or strength gains³⁷. Furthermore, the American Heart Association (AHA)³⁸ recommends that the initial weight used for resistance training in the elderly be approximately 30-40%1RM for one set of 10-15 repetitions two days per week which may be inadequate to promote gains in muscle mass, due to the importance of exercise volume, particularly with the muscle protein synthesis (MPS) response³⁹.

Blood flow restricted exercise, also known as occlusion training, offers a novel mode of exercise for the elderly, as it allows the achievement of skeletal muscle hypertrophy at low intensities. Occlusion training involves applying a wrapping device, typically a modified blood pressure cuff, proximal to the muscle being trained⁴⁰. Research indicates that occlusion training can provide a unique, beneficial mode of exercise in clinical settings, as it produces positive training adaptations at intensities equivalent to physical activities of daily life (10-30% of maximal work capacity)⁴¹. Numerous studies have shown that intensities as low as 20% 1RM with moderate blood flow restriction (100mmHG) can lead to significant increases in skeletal muscle hypertrophy⁴¹⁻⁴⁵, strength^{34,41-47}, and endurance^{43,46}.

Occlusion training works through a variety of mechanisms including the recruitment of fast twitch fibers at low intensities and the stimulation of the protein synthe-

sis pathway mammalian target of rapamycin (mTOR)⁴⁸. Additionally, this type of training has demonstrated growth hormone (GH) responses 290 times above baseline, which is higher than that seen with traditional high intensity exercise⁴⁹.

Although a majority of the research has been completed in young healthy subjects, a few studies have demonstrated favorable outcomes with those of advancing age. Takarada et al³⁴ investigated elbow flexion exercise twice a week for 16 weeks (~50% 1RM) with low intensity exercise (~50% 1RM) with and without occlusion and heavy resistance training (~80% 1RM) in older women (~58 years). Interestingly, low intensity exercise with occlusion increased CSA of the elbow flexors and extensors, as well as isokinetic strength significantly more than low intensity exercise. Isokinetic strength gains were similar to those seen in the high intensity exercise group.

Karabulut et al⁴⁷ investigated the effects of occlusion training in older males (50-64 years). A high intensity group (80%1RM) and a low intensity group with and without occlusion (20%1RM) performed three upper body and two lower body exercises three times a week for 6 weeks. They found that the high intensity group and the low intensity group with occlusion had significantly greater strength increases in every exercise when compared with low intensity exercise. However, they did note that the percentage increase in the high intensity groups leg extension strength was significantly greater than low intensity exercise with and without occlusion. The researchers concluded that low intensity occlusion training was almost as effective as high intensity training in older males.

Fry et al⁵⁰ reports perhaps the most interesting acute study to date with low intensity occlusion training in the elderly (~70 years). This groups of researchers found that bilateral knee extensor exercise at low intensities (~20% 1RM) with occlusion was able to significantly stimulate protein synthesis. This is of great importance, because as previously stated, many elderly are unable to train at higher intensities, and this study demonstrated that low intensity exercise without occlusion does not increase protein synthesis. Thus, low intensity exercise without occlusion would likely not produce a stimulus strong enough to promote muscle hypertrophy or maintain muscle mass. Additionally they monitored for the risk of deep vein thrombosis by measuring D-dimer (marker of intravascular clot formation), which was unaffected with either condition.

In conclusion, numerous studies show the benefits of occlusion training with respect to skeletal muscle hypertrophy, strength, and endurance. In addition, the few studies completed on the elderly with occlusion showed favorable outcomes similar to that of higher intensity exercise. It should be noted that adaptation occurs without significant elevations of any known markers for muscle damage (myoglobin, lipid peroxide, creatine kinase)^{41,49}.

Nutrition Intervention

In addition to training, proper diet with respect to dietary protein is essential in preserving skeletal muscle

mass. With aging comes a decline in the bodies sensitivity to amino acids and numerous studies suggest that the protein requirement for the elderly is elevated compared to the young⁵¹⁻⁵³. The United States recommended daily allowance (RDA) for protein is .8 g/kg/day which is the amount of protein necessary to sustain life, not optimal function. Newer research suggests that we should focus not on the overall daily amount of dietary protein, but instead the optimal dose needed to maximally stimulate MPS and the optimal amount of time between meals until we can maximally stimulate MPS again^{54, 55}.

It is generally accepted that 25-30g of high quality protein, is the amount needed per meal to maximally stimulate MPS in the elderly^{56, 57}. To illustrate, Symons et al⁵⁸ found that 60g of protein stimulated MPS to no greater degree than 30g per meal. This phenomenon is referred to as refractory; meaning ~30g of protein appears to be the ceiling for the anabolic response to a meal.

In addition to the protein dose at a meal, Norton et al⁵⁴ investigated the duration of MPS from a complete meal in which they showed MPS was elevated for three hours before returning to baseline. Interestingly, although MPS had returned back to baseline, plasma leucine concentrations and the initiation factors for MPS remained elevated. They concluded that eating another meal three hours following the previous meal would likely fail to elevate MPS because the initiation factors for MPS were still elevated⁵⁵. Infusion research supports these findings as Bohe et al⁵⁹ found that MPS returned to baseline after two hours despite purified essential amino acids (EAA) being infused for eight hours. Norton and Wilson⁵⁵ suggest that amino acid levels should be spiked with a meal, then given time to fall back towards baseline, followed by another amino acid spike 4-5 hours later. This strategy seems especially important in aging, when optimal nutrition for skeletal muscle maintenance is of absolute importance. Unfortunately, most people regardless of age inadequately distribute their protein throughout the day, consuming over 65% of their daily protein after 1830 h⁶⁰. Emphasis should be placed upon equal distribution of protein, resulting in more maximal protein stimulations throughout the day, theoretically improving skeletal muscle health.

In conclusion, adequate distribution of protein throughout the day is important to maximize muscle anabolism. The elderly should focus on consuming at least 25g of protein three times a day, separated by approximately 4-5 hours. Eating at 800h, 1300h, and 1800h would maximally stimulate protein synthesis three times a day promoting skeletal muscle hypertrophy and/or maintenance of muscle mass.

Resistance Training and Nutrition Interventions

Research has shown that when resistance training is combined with a nutritional intervention, benefits in both strength and body composition are observed. Campbell and colleagues provide the most convincing evidence that emphasis be placed not only on resistance training

but also dietary protein⁶¹. They compiled data over 15 years from 106 men and women between the ages of 50 and 80 years who had participated in one of their diet and resistance training studies. The training programs they used for all studies were similar in duration, intensity, exercises, frequency and equipment used. In addition, the dietary protein intakes ranged from .4g/kg to 1.7g/kg. Upon compiling the data they found an apparent loss of fat-free mass by many of their older subjects when they consumed the RDA for protein (0.8g/kg), despite the anabolic stimulus from the resistance training bout. They concluded from their analysis that an older subject who consumed the RDA for protein would theoretically lose about 0.2 kg of fat-free mass after a 12-week resistance training program and protein intakes of 1.0, 1.2, 1.4, and 1.6g/kg would translate into fat free mass increases of 0.0, 0.3, 0.5, and 0.8 kg, respectively⁶¹.

These findings highlight an importance of not only resistance training but also adequate dietary protein distributed throughout the day. It should be noted that while occlusion training has resulted in significant elevations in protein synthesis, no study has investigated this stimulus in combination with a nutritional intervention; however the authors hypothesize the response to be similar to that of higher intensity exercise.

Drug Intervention

As with any disease or condition, numerous drugs are developed in hopes of combating the negative effects of the condition and to improve clinical outcomes. With aging, circulating levels of numerous muscle anabolic hormones decline; such as testosterone, GH and insulin-like growth factor-one (IGF-1)⁶². The research thus far is conflicting; however it does appear that exercise, regardless of the drug employed, is necessary for the greatest gain in strength and muscle mass.

In males, bioavailable testosterone levels decline by as much as 64% between the ages of 25 and 85, while women drop only 28%^{63, 64}. The reasoning behind testosterone administration is that satellite cells decrease with sarcopenia and when testosterone is given there appears to be an increase of satellite cells in a dose dependent manner^{65, 66}. Several studies have shown a direct relationship between serum testosterone and muscle strength in older men, however this relationship does not hold true for women^{9, 67, 68}. Ryall et al⁶² believe the disparity lies with the more than twofold increase in sex hormone-binding globulin (SHBG) over a lifespan in males, which remains unchanged in females. SHBG functions include binding anabolic hormones, which could possibly account for decrease in strength observed in males but not females.

Circulating GH is another anabolic hormone that progressively declines after age 30 at an average rate greater than 1% per year^{69, 70}. Although GH exerts many actions through its production of IGF-1, GH is capable of promoting skeletal muscle growth independent of

IGF-1⁷¹. Despite the requirement for endogenous GH to maintain muscle and bone, exogenous GH does not

increase functional (no concomitant strength increase) fat free mass (FFM)⁷². The increased FFM is attributed to fluid retention or an increase in collagen⁷³. The lack of benefit seen with exogenous GH administration is attributed to the complexity of the GH/IGF-1 pathway itself and the numerous GH isoforms; all of which contribute to the difficulty of synthetically mimicking the effects of endogenous GH secretion. It is conceivable that the endogenous response from low intensity blood flow restricted exercise could potentially play a more important role than the synthetic form⁴⁹.

Recently, work has focused on the effects of IGF-1 administration combined with its predominant circulating binding protein IGFBP-3 on the elderly⁷⁴. Boonen et al⁷⁵ reported that the use of this combination allows for a higher dose of IGF-1, without the negative effects of hypoglycemia noted with IGF-1 alone. The combination also preserved femoral bone mass and increased grip strength in elderly women. While the results of this study are promising drawing any definitive conclusions is difficult due to a small sample size.

Drug administration may have some protective role with the aging population to help maintain muscle mass and strength, but exogenous administration of those hormones has produced confounding outcomes. Proper exercise and diet likely play a more prominent role in the preservation of muscle mass and strength than administration of synthetic hormones, without the potential negative side effects of drug use.

Conclusion

The actual cause of sarcopenia is unknown but appears multifactorial. Interventions should focus on resistance training and adequate nutrition. A novel training stimulus such as occlusion training might make resistance training more applicable to the elderly, because this mode of training involves a low-load, low-intensity bout of exercise. Furthermore, an emphasis must be placed on high quality protein adequately distributed throughout the day to maximize protein synthesis. Additionally, drug therapy might offer some help but the research remains equivocal. In summation, proper training and nutrition might not be able to completely stop sarcopenia, but it may attenuate its progression.

References

- Koopman R, van Loon LJ. Aging, exercise, and muscle protein metabolism. *J Appl Physiol*. 2009; 106: 2040-2048.
- Mourtzakis M, Bedbrook M. Muscle atrophy in cancer: a role for nutrition and exercise. *Appl Physiol Nutr Metab*. 2009; 34: 950-956.
- Holloszy JO. Exercise-induced increase in muscle insulin sensitivity. *J Appl Physiol*. 2005; 99: 338-343.
- Helge JW, Biba TO, Galbo H, Gaster M, Donsmark M. Muscle triacylglycerol and hormone-sensitive lipase activity in untrained and trained human muscles. *Eur J Appl Physiol*. 2006; 97: 566-572.
- Sahlin K, Mogensen M, Bagger M, Fernstrom M, Pedersen PK. The potential for mitochondrial fat oxidation in human skeletal muscle influences whole body fat oxidation during low-intensity exercise. *Am J Physiol Endocrinol Metab*. 2007; 292: E223-230.
- Bosy-Westphal A, Reinecke U, Schlorke T, Illner K, Kutzner D, Heller M, et al. Effect of organ and tissue masses on resting energy expenditure in underweight, normal weight and obese adults. *Int J Obes Relat Metab Disord*. 2004; 28: 72-79.
- Frontera WR, Hughes VA, Fielding RA, Fiatarone MA, Evans WJ, Roubenoff R. Aging of skeletal muscle: a 12-yr longitudinal study. *J Appl Physiol*. 2000; 88: 1321-1326.
- Frontera WR, Hughes VA, Lutz KJ, Evans WJ. A cross-sectional study of muscle strength and mass in 45- to 78-yr-old men and women. *J Appl Physiol*. 1991; 71: 644-650.
- Iannuzzi-Sucich M, Prestwood KM, Kenny AM. Prevalence of sarcopenia and predictors of skeletal muscle mass in healthy, older men and women. *J Gerontol A Biol Sci Med Sci*. 2002; 57: M772-777.
- Kenny AM, Dawson L, Kleppinger A, Iannuzzi-Sucich M, Judge JO. Prevalence of sarcopenia and predictors of skeletal muscle mass in nonobese women who are long-term users of estrogen-replacement therapy. *J Gerontol A Biol Sci Med Sci*. 2003; 58: M436-440.
- Rolland Y, Lauwers-Cances V, Cournot M, Nourhashemi F, Reynish W, Riviere D, et al. Sarcopenia, calf circumference, and physical function of elderly women: a cross-sectional study. *J Am Geriatr Soc*. 2003; 51: 1120-1124.
- Chan BK, Marshall LM, Winters KM, Faulkner KA, Schwartz AV, Orwoll ES. Incident fall risk and physical activity and physical performance among older men: the Osteoporotic Fractures in Men Study. *Am J Epidemiol*. 2007; 165: 696-703.
- Orr R, de Vos NJ, Singh NA, Ross DA, Stavrinou TM, Fiatarone-Singh MA. Power training improves balance in healthy older adults. *J Gerontol A Biol Sci Med Sci*. 2006; 61: 78-85.
- Metter EJ, Talbot LA, Schrager M, Conwit R. Skeletal muscle strength as a predictor of all-cause mortality in healthy men. *J Gerontol A Biol Sci Med Sci*. 2002; 57: B359-365.
- Doherty TJ, Brown WF. The estimated numbers and relative sizes of thenar motor units as selected by multiple point stimulation in young and older adults. *Muscle Nerve*. 1993; 16: 355-366.
- Doherty TJ, Brown WF. Age-related changes in the twitch contractile properties of human thenar motor units. *J Appl Physiol*. 1997; 82: 93-101.
- Roos MR, Rice CL, Vandervoort AA. Age-related changes in motor unit function. *Muscle Nerve*. 1997; 20: 679-690.
- Kosek DJ, Kim JS, Petrella JK, Cross JM, Bamman MM. Efficacy of 3 days/wk resistance training on myofiber hypertrophy and myogenic mechanisms in young vs. older adults. *J Appl Physiol*. 2006; 101: 531-544.
- Welle S, Thornton C, Statt M. Myofibrillar protein synthesis in young and old human subjects after three months of resistance training. *Am J Physiol*. 1995; 268: E422-427.
- Dangin M, Boirie Y, Guillet C, Beaufrere B. Influence of the protein digestion rate on protein turnover in young and elderly subjects. *J Nutr*. 2002; 132: 3228S-3233S.
- Dangin M, Guillet C, Garcia-Rodenas C, Gachon P, Bouteloup-Demange C, Reiffers-Magnani K, et al. The rate of protein digestion affects protein gain differently during aging in humans. *J Physiol*. 2003; 549: 635-644.
- Katsanos CS, Kobayashi H, Sheffield-Moore M, Aarsland A, Wolfe RR. A high proportion of leucine is required for optimal stimulation of the rate of muscle protein synthesis by essential amino acids in the elderly. *Am J Physiol Endocrinol Metab*. 2006; 291: E381-387.
- Morley JE. Anorexia, sarcopenia, and aging. *Nutrition*. 2001; 17: 660-663.
- Murton AJ, Constantin D, Greenhaff PL. The involvement of the ubiquitin proteasome system in human skeletal muscle remodeling and atrophy. *Biochim Biophys Acta*. 2008; 1782: 730-743.

25. Greenhaff PL, Karagounis LG, Peirce N, Simpson EJ, Hazell M, Layfield R, et al. Disassociation between the effects of amino acids and insulin on signaling, ubiquitin ligases, and protein turnover in human muscle. *Am J Physiol Endocrinol Metab.* 2008; 295: E595-604.
26. Guillet C, Auguste P, Mayo W, Kreher P, Gascan H. Ciliary neurotrophic factor is a regulator of muscular strength in aging. *J Neurosci.* 1999; 19: 1257-1262.
27. Delbono O. Neural control of aging skeletal muscle. *Aging Cell.* 2003; 2: 21-29.
28. Narici MV, Maffulli N. Sarcopenia: characteristics, mechanisms and functional significance. *Br Med Bull.* 2010; 95: 139-159.
29. Dirks AJ, Leeuwenburgh C. The role of apoptosis in age-related skeletal muscle atrophy. *Sports Med.* 2005; 35: 473-483.
30. Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, Bruder CE, et al. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature.* 2004; 429: 417-423.
31. Brose A, Parise G, Tarnopolsky MA. Creatine supplementation enhances isometric strength and body composition improvements following strength exercise training in older adults. *J Gerontol A Biol Sci Med Sci.* 2003; 58: 11-19.
32. Ferri A, Scaglioni G, Pousson M, Capodaglio P, Van Hoecke J, Narici MV. Strength and power changes of the human plantar flexors and knee extensors in response to resistance training in old age. *Acta Physiol Scand.* 2003; 177: 69-78.
33. Frontera WR, Hughes VA, Krivickas LS, Kim SK, Foldvari M, Roubenoff R. Strength training in older women: early and late changes in whole muscle and single cells. *Muscle Nerve.* 2003; 28: 601-608.
34. Takarada Y, Takazawa H, Sato Y, Takebayashi S, Tanaka Y, Ishii N. Effects of resistance exercise combined with moderate vascular occlusion on muscular function in humans. *J Appl Physiol.* 2000; 88: 2097-2106.
35. Fujita S, Abe T, Drummond MJ, Cadenas JG, Dreyer HC, Sato Y, et al. Blood flow restriction during low-intensity resistance exercise increases S6K1 phosphorylation and muscle protein synthesis. *J Appl Physiol.* 2007; 103: 903-910.
36. Lexell J. Ageing and human muscle: observations from Sweden. *Can J Appl Physiol.* 1993; 18: 2-18.
37. ACSM. American College of Sports Medicine position stand. Progression models in resistance training for healthy adults. *Med Sci Sports Exerc.* 2009; 41: 687-708.
38. Williams MA, Haskell WL, Ades PA, Amsterdam EA, Bittner V, Franklin BA, et al. Resistance exercise in individuals with and without cardiovascular disease: 2007 update: a scientific statement from the American Heart Association Council on Clinical Cardiology and Council on Nutrition, Physical Activity, and Metabolism. *Circulation.* 2007; 116: 572-584.
39. Burd NA, Holwerda AM, Selby KC, West DW, Staples AW, Cain NE, et al. Resistance exercise volume affects myofibrillar protein synthesis and anabolic signalling molecule phosphorylation in young men. *J Physiol.* 2010; 588: 3119-3130.
40. Loenneke JP, Pujol TJ. The Use of Occlusion Training to Produce Muscle Hypertrophy. *Strength Cond J.* 2009; 31: 77-84.
41. Abe T, Kearns CF, Sato Y. Muscle size and strength are increased following walk training with restricted venous blood flow from the leg muscle, Kaatsu-walk training. *J Appl Physiol.* 2006; 100: 1460-1466.
42. Madarame H, Neya M, Ochi E, Nakazato K, Sato Y, Ishii N. Cross-transfer effects of resistance training with blood flow restriction. *Med Sci Sports Exerc.* 2008; 40: 258-263.
43. Takarada Y, Sato Y, Ishii N. Effects of resistance exercise combined with vascular occlusion on muscle function in athletes. *Eur J Appl Physiol.* 2002; 86: 308-314.
44. Takarada Y, Tsuruta T, Ishii N. Cooperative effects of exercise and occlusive stimuli on muscular function in low-intensity resistance exercise with moderate vascular occlusion. *Jpn J Physiol.* 2004; 54: 585-592.
45. Yasuda T, Fujita S, Ogasawara R, Sato Y, Abe T. Effects of low-intensity bench press training with restricted arm muscle blood flow on chest muscle hypertrophy: a pilot study. *Clin Physiol Funct Imaging.* 2010.
46. Sumide T, Sakuraba K, Sawaki K, Ohmura H, Tamura Y. Effect of resistance exercise training combined with relatively low vascular occlusion. *J Sci Med Sport.* 2009; 12: 107-112.
47. Karabulut M, Abe T, Sato Y, Bembem MG. The effects of low-intensity resistance training with vascular restriction on leg muscle strength in older men. *Eur J Appl Physiol.* 2010; 108: 147-155.
48. Loenneke JP, Wilson GJ, Wilson JM. A Mechanistic Approach to Blood Flow Occlusion. *Int J Sports Med.* 2010; 31: 1-4.
49. Takarada Y, Nakamura Y, Aruga S, Onda T, Miyazaki S, Ishii N. Rapid increase in plasma growth hormone after low-intensity resistance exercise with vascular occlusion. *J Appl Physiol.* 2000; 88: 61-65.
50. Fry CS, Glynn EL, Drummond MJ, Timmerman KL, Fujita S, Abe T, et al. Blood flow restriction exercise stimulates mTORC1 signaling and muscle protein synthesis in older men. *J Appl Physiol.* 2010; 108: 1199-1209.
51. Houston DK, Nicklas BJ, Ding J, Harris TB, Tyllavsky FA, Newman AB, et al. Dietary protein intake is associated with lean mass change in older, community-dwelling adults: the Health, Aging, and Body Composition (Health ABC) Study. *Am J Clin Nutr.* 2008; 87: 150-155.
52. Peppersack T, Corrette M, Beyer I, Namias B, Andr S, Benoit F, et al. Examining the effect of intervention to nutritional problems of hospitalised elderly: a pilot project. *J Nutr Health Aging.* 2002; 6: 306-310.
53. Wolfe RR, Miller SL. The recommended dietary allowance of protein: a misunderstood concept. *JAMA.* 2008; 299: 2891-2893.
54. Norton LE, Layman DK, Bunpo P, Anthony TG, Brana DV, Garlick PJ. The leucine content of a complete meal directs peak activation but not duration of skeletal muscle protein synthesis and mammalian target of rapamycin signaling in rats. *J Nutr.* 2009; 139: 1103-1109.
55. Norton LE, Wilson GJ. Optimal protein intake to maximize muscle protein synthesis. Examinations of optimal meal protein intake and frequency for athletes. *Agro Food Ind High-Tech.* 2009; 20: 54-57.
56. Millward DJ, Layman DK, Tome D, Schaafsma G. Protein quality assessment: impact of expanding understanding of protein and amino acid needs for optimal health. *Am J Clin Nutr.* 2008; 87: 1576S-1581S.
57. Paddon-Jones D, Sheffield-Moore M, Zhang XJ, Volpi E, Wolf SE, Aarsland A, et al. Amino acid ingestion improves muscle protein synthesis in the young and elderly. *Am J Physiol Endocrinol Metab.* 2004; 286: E321-328.
58. Symons TB, Sheffield-Moore M, Wolfe RR, Paddon-Jones D. A moderate serving of high-quality protein maximally stimulates skeletal muscle protein synthesis in young and elderly subjects. *J Am Diet Assoc.* 2009; 109: 1582-1586.
59. Bohe J, Low JF, Wolfe RR, Rennie MJ. Latency and duration of stimulation of human muscle protein synthesis during continuous infusion of amino acids. *J Physiol.* 2001; 532: 575-579.
60. de Castro JM. Circadian rhythms of the spontaneous meal pattern, macronutrient intake, and mood of humans. *Physiol Behav.* 1987; 40: 437-446.
61. Campbell WW, Leidy HJ. Dietary protein and resistance training effects on muscle and body composition in older persons. *J Am Coll Nutr.* 2007; 26: 696S-703S.
62. Ryall JG, Schertzer JD, Lynch GS. Cellular and molecular mechanisms underlying age-related skeletal muscle wasting and weakness. *Biogerontology.* 2008; 9: 213-228.

63. Khosla S, Melton LJ, 3rd, Atkinson EJ, O'Fallon WM, Klee GG, Riggs BL. Relationship of serum sex steroid levels and bone turnover markers with bone mineral density in men and women: a key role for bioavailable estrogen. *J Clin Endocrinol Metab.* 1998; 83: 2266-2274.
64. Morley JE, Kaiser F, Raum WJ, Perry HM, 3rd, Flood JF, Jensen J, et al. Potentially predictive and manipulable blood serum correlates of aging in the healthy human male: progressive decreases in bioavailable testosterone, dehydroepiandrosterone sulfate, and the ratio of insulin-like growth factor 1 to growth hormone. *Proc Natl Acad Sci U S A.* 1997; 94: 7537-7542.
65. Sinha-Hikim I, Cornford M, Gaytan H, Lee ML, Bhasin S. Effects of testosterone supplementation on skeletal muscle fiber hypertrophy and satellite cells in community-dwelling older men. *J Clin Endocrinol Metab.* 2006; 91: 3024-3033.
66. Sinha-Hikim I, Taylor WE, Gonzalez-Cadavid NF, Zheng W, Bhasin S. Androgen receptor in human skeletal muscle and cultured muscle satellite cells: up-regulation by androgen treatment. *J Clin Endocrinol Metab.* 2004; 89: 5245-5255.
67. Schaap LA, Pluijm SM, Smit JH, van Schoor NM, Visser M, Gooren LJ, et al. The association of sex hormone levels with poor mobility, low muscle strength and incidence of falls among older men and women. *Clin Endocrinol (Oxf).* 2005; 63: 152-160.
68. Waters DL, Yau CL, Montoya GD, Baumgartner RN. Serum Sex Hormones, IGF-1, and IGFBP3 Exert a Sexually Dimorphic Effect on Lean Body Mass in Aging. *J Gerontol A Biol Sci Med Sci.* 2003; 58: 648-652.
69. Hermann M, Berger P. Hormonal changes in aging men: a therapeutic indication? *Exp Gerontol.* 2001; 36: 1075-1082.
70. Zadik Z, Chalew SA, McCarter RJ, Jr., Meistas M, Kowarski AA. The influence of age on the 24-hour integrated concentration of growth hormone in normal individuals. *J Clin Endocrinol Metab.* 1985; 60: 513-516.
71. Sotiropoulos A, Ohanna M, Kedzia C, Menon RK, Kopchick JJ, Kelly PA, et al. Growth hormone promotes skeletal muscle cell fusion independent of insulin-like growth factor 1 up-regulation. *Proc Natl Acad Sci U S A.* 2006; 103: 7315-7320.
72. Papadakis MA, Grady D, Black D, Tierney MJ, Gooding GA, Schambelan M, et al. Growth hormone replacement in healthy older men improves body composition but not functional ability. *Ann Intern Med.* 1996; 124: 708-716.
73. Ehrnborg C, Rosen T. Physiological and pharmacological basis for the ergogenic effects of growth hormone in elite sports. *Asian J Androl.* 2008; 10: 373-383.
74. Borst SE. Interventions for sarcopenia and muscle weakness in older people. *Age Ageing.* 2004; 33: 548-555.
75. Boonen S, Rosen C, Bouillon R, Sommer A, McKay M, Rosen D, et al. Musculoskeletal effects of the recombinant human IGF-I/IGF binding protein-3 complex in osteoporotic patients with proximal femoral fracture: a double-blind, placebo-controlled pilot study. *J Clin Endocrinol Metab.* 2002; 87: 1593-1599.