

Genetics of Polycystic Ovary Syndrome

Prapas N, Karkanaki A, Prapas I, Kalogiannidis I, Katsikis I, Panidis D

Division of Endocrinology and Human Reproduction, 2nd Department of Obstetrics and Gynecology, Aristotle University of Thessaloniki, Greece

Abstract

Polycystic ovary syndrome (PCOS) is a syndrome involving defects in primary cellular control mechanisms that result in the expression of chronic anovulation and hyperandrogenism. This syndrome has been for many years one of the most controversial entities in gynecological endocrinology. Polycystic ovary syndrome has been proven to be a familial condition. Although the role of genetic factors in PCOS is strongly supported, the genes that are involved in the etiology of the syndrome have not been fully investigated until now, as well as the environmental contribution in their expression. The heterogeneity of the syndrome entertains the mystery around this condition which concerns thousands of infertile women worldwide. Some genes have shown altered expression suggesting that the genetic abnormality in PCOS affects signal transduction pathways controlling steroidogenesis, steroid hormones action, gonadotrophin action and regulation, insulin action and secretion, energy homeostasis, chronic inflammation and others. The present review of the contemporary literature constitutes an effort to present all the trends in the current research for the etiology of polycystic ovary syndrome. Hippokratia 2009; 13 (4): 216-223

Key words: polycystic ovary syndrome, etiology of PCOS genetics of PCOS

Corresponding author: Karkanaki A, 29B, V Olgas Str, 52422, Thessaloniki, Greece, tel. 6932 315022, e-mail address: artemisk@med.auth.gr

Polycystic Ovary Syndrome (PCOS) is a familial condition, as has long been noted^{1,2}. Some clinical genetic studies have pointed to an autosomal dominant inheritance^{1,3,4} while others showed that it was more likely that the syndrome is a complex trait with oligogenic basis^{5,6}. Although clustering of cases in families strongly support the role of genetic factors in the development of PCOS, heterogeneity of phenotypic features in different families and even within the same family underscores the importance of the environmental contribution. Modifications of molecular structure of gonadotrophins, their receptors and of the enzymes involved in steroidogenesis, insulin action and secretion have been under continuous and intense investigation with variable results. Whereas several positive results have been reported, there are no genes universally accepted fundamentally important in PCOS aetiology. This has resulted partially because of various factors such as the lack of a worldwide accepted diagnostic scheme for PCOS, diagnostic capability only in reproductive-aged women, limited number of patients in case-control studies, analysis of only one or two variants of candidate genes and incomplete knowledge of pathophysiology of the syndrome.

Two possible approaches are used to identify a genetic locus for PCOS genes: (i) association studies where a predisposing allele is expected to be found more frequently in the affected population than the normal individuals and (ii) linkage studies where the probands and their families are investigated to determine if particular genomic landmarks are distributed independently or in

linkage with the phenotype. While the mode of inheritance is not required for the association studies, it requires a relatively large set of individuals for a clear conclusion⁷. Many genes presented altered expression suggesting thus that the genetic abnormality in PCOS affects signal transduction ruling steroidogenesis, steroid hormones action, gonadotrophin action and regulation, insulin action and secretion, energy homeostasis, chronic inflammation and others.

Genes involved in ovarian and adrenal steroidogenesis

The first step in steroidogenesis is the conversion of cholesterol into progesterone, catalyzed by the P450 cytochrome side chain cleavage enzyme encoded by CYP11a gene located at 15q24⁸. Investigation of CYP11A gene showed a significant association between serum testosterone levels and the alleles of the CYP11a with a 5' untranslated region (UTR) consisting of repeats of a (ttta)n pentanucleotide, a variable number tandem repeat (VNTR) polymorphism⁹. Two other case-control studies^{10,11}, confirmed these findings in support of the encouraging evidence for the association between CYP11a and PCOS. However, subsequent studies^{12,13} have failed to find any significant association between this gene locus and its VNTR alleles and PCOS. Further investigation is required due to these controversial results in order to confirm a role in the aetiology of PCOS of this gene.

Another part in steroidogenesis is the conversion of 17-hydroxyprogesterone into 11-deoxycortisol which

is catalyzed by the 21-hydroxylase enzyme encoded by CYP21. The deficiency of this enzyme is responsible for most cases of congenital adrenal hyperplasia and increased serum 17-hydroxyprogesterone levels are correlated with its deficiency. It is a common finding among women with functional hyperandrogenism or PCOS an increased serum 17-hydroxyprogesterone response to ACTH stimulation^{14,15}. Furthermore, patients having both heterozygote CYP21 mutations and clinical symptoms exhibit a PCOS-like phenotype¹⁶. Accordingly, mutations of CYP21 have been investigated as a candidate gene in patients with PCOS. Two studies showed that children with premature pubarche and adolescent girls with hyperandrogenism were heterozygous for mutations in CYP21^{16,17}. On the other hand, there are other researchers that found no clear concordance between the CYP21 genotype and the functional origin of androgen excess^{18,19}. Overall, CYP21 and associated mutations do not seem to play a key role in the development of PCOS.

The conversion of pregnenolone and progesterone into 17-hydroxypregnenolone and 17-hydroxyprogesterone, respectively, and of these steroids into dehydroepiandrosterone (DHEA) and Δ_4 -Androstendione (Δ_4 A) is catalyzed by the P450c17 α enzyme. This enzyme has both 17 α -hydroxylase and 17,20-lyase activities and is encoded by CYP17 located at 10q24.3²⁰. It was reported increased P450c17 α expression and enzymatic activity in ovarian theca cells from women with PCOS as well as increased transactivation of the CYP17 promoter²¹⁻²³. Moreover, it was showed that CYP17 expression is dysregulated at the level of mRNA stability in PCOS theca cells²⁴. Another study identified a rare T/C single nucleotide polymorphism (SNP) in the promoter region of CYP17 increasing the susceptibility to develop PCOS²⁵. Subsequently, more comprehensive studies have failed to detect a significant linkage between CYP17 and PCOS²⁶⁻²⁹. Although CYP17 gene does not seem to be a candidate gene in the pathophysiology of PCOS, it should be noted that post-translational regulation of this gene product might play a role in the pathophysiology of PCOS⁷.

The enzyme complex aromatase converts androgens to estrogens. This enzyme complex is composed of the cytochrome P450 aromatase and the NADPH cytochrome P450 reductase³⁰, and P450arom is encoded by CYP19 located at 15p21.1³¹. Aromatase deficiency has been reported in a number of hyperandrogenic patients^{32,33}. It has been demonstrated that granulosa cells obtained from medium-sized follicles of women with PCOS have little aromatase activity³⁴. Similarly, it has been showed that when compared to the control follicles, all PCOS follicles contained low levels of P450arom mRNA, estradiol, and lower aromatase stimulating bioactivity³⁵. These findings indicate that the aromatase activity might be decreased in PCOS follicles, and that the possible androgen excess resulting might contribute to abnormal follicle development. Association studies utilizing SNPs and haplotypes showed association with PCOS symptoms and serum testosterone levels^{36,37}.

Genes involved in steroid hormone actions

All androgens transmit their signal through the androgen receptor which belongs to a family of nuclear transcription factors. The androgen receptor is encoded by the gene (AR) located at Xq11-12³⁸ and is composed of three functional domains: the transactivation domain, the DNA binding domain, and the ligand-binding domain. A VNTR polymorphism consisting of CAG repeats in exon-1 encoding a polyglutamine chain in the N-terminal transactivation domain is embedded in AR³⁹. The transcriptional activity of androgen receptor is inversely correlated with the number of CAG repeats⁴⁰. Variations of these repeats, even within the normal polymorphic range, have been related to various disorders associated with low- or high-androgenic activities⁴¹⁻⁴³. Therefore, decreased number of CAG repeats with an increased androgen receptor activity could explain some of the PCOS phenotype exhibiting the normal serum androgen levels and hyperandrogenism symptoms⁴⁴. Nevertheless, some studies failed to prove any association between this VNTR and PCOS^{44,45}. On the contrary, other studies demonstrated a significantly greater frequency of alleles with longer CAG repeats for infertile PCOS patients compared with fertile women⁴⁶. Concluding, AR gene is not a strong candidate for the etiology of PCOS.

Serum Sex Hormone-Binding Globulin (SHBG) levels are commonly low in patients with hyperandrogenism, especially in association with PCOS⁴⁷. SHBG is composed of a homodimeric glycoprotein produced by hepatocytes and is encoded by a 4-kb gene at the 17p12-p13^{48,49}. A pentanucleotide repeat polymorphism, at the promoter of SHBG gene has been described to influence the transcriptional activity of SHBG gene⁵⁰. Consequently, it has been investigated whether this polymorphism is associated with PCOS and whether polymorphic variants of the gene are related to serum SHBG levels in women with PCOS⁵¹. A significant association was found between this polymorphism and PCOS⁵¹. PCOS patients carrying the longer allele genotypes had lower SHBG levels. In accordance with the latter result, Cousin et al.⁵² recently demonstrated that longer alleles lowered serum SHBG levels in hirsute women when compared with six repeat alleles. Although Urbanek et al.⁴⁵ did not find any association or linkage between a marker close to the locus and PCOS, it could be concluded that SHBG gene is a potential candidate gene in the pathogenesis of PCOS⁷.

Genes involved in gonadotropin action and regulation

The gene encoding the β -subunit of LH which is responsible for LH specificity, has been explored in PCOS patients⁷. Initially, it was identified an abnormal form of LH with two-point mutations, Trp8Arg and Ile15Thr, in the LH β -subunit gene⁵³. In addition, these mutations produced structural changes in the variant LH molecules (v-LH)⁵⁴ and caused v-LH to have an increased in vitro activity and a decreased in vivo half life compared to that of non mutant form⁵⁵. However, in vivo activity of v-LH

could not be explained. The implication of v-LH in both healthy women and PCOS patients was explored and it was found that the occurrence of these mutations in LH β -subunit gene was not higher in PCOS compared with healthy women^{7,56}. On the other hand, subgroup analysis of this study revealed that obese PCOS patients had a higher frequency of the heterozygous v-LH compared with obese controls^{7,56}. However, other studies failed to find any association with PCOS⁵⁷⁻⁵⁹. The assumption that an activating mutation in the LH receptor gene could trigger hyperandrogenism in patients with PCOS having normal serum LH concentrations and high androgen levels was demolished⁸. Overall, the functional role of the v-LHs is unclear but it seems not to be crucial in PCOS pathogenesis or female infertility.

Follistatin, a monomeric glycoprotein encoded by a single gene, is linked functionally through its role as a high-affinity binding protein for activin⁶⁰. Activin is dimeric glycoprotein which belongs to the TGF- β superfamily, induces FSH and insulin secretion, ovarian follicular maturation and inhibits LH-stimulated ovarian androgen production⁶⁰. Actually, overexpression of follistatin in transgenic mice resulted in suppression of serum levels of FSH and arrested ovarian folliculogenesis⁶¹. Therefore overwhelming activin neutralization due to increased follistatin reduces FSH concentrations, arrests follicular maturation, augments androgen production, and impairs insulin release. Because all of these changes are typical features of PCOS⁶², follistatin gene has been explored as a candidate gene in PCOS. The results of distinct studies are conflicting and significant linkage was failed to be proven^{45,63,64}.

Genes involved in insulin action and secretion

The insulin gene (INS) is located between the genes for tyrosine hydroxylase and for IGF-II at 11p15.5, and includes variable tandem repeats (VNTR) embedded at the 5' regulatory region of INS⁶⁵. The VNTR polymorphism regulates the transcriptional rate of the INS⁶⁶ and probably that of the gene encoding IGF-II⁶⁷. The number of the repeats of the INS VNTR ranges from 26 to 200, and due to this feature INS VNTR polymorphism has three size classes. Class-I alleles comprise the shorter polymorphism, consisting of a length of 40 repeats. Class-II alleles are composed usually of 80 repeats and are uncommon in Caucasian. Class-III alleles compose the longest polymorphic region having an average of 157 repeats⁶⁸. Transcriptional activity of the longer polymorphic region is greater than that of the shorter one⁶⁶. Besides their effect on regulating INS expression, they have been implicated in the pathogenesis of type-2 Diabetes Mellitus (DM) in many studies^{69,70}. The hyperinsulinemia in PCOS may be the result of primary insulin resistance or the direct effect of pancreatic β -cell disorder as defects in both insulin action^{71, 72} and in pancreatic β -cell function^{73,74} have been reported. Therefore, it was evaluated the linkage and association of the INS VNTR polymorphisms in families with affected members with PCOS⁷⁵.

An association was found between PCOS and allelic variation at the INS VNTR locus in three separate populations⁷⁵. Furthermore, it was found that class III alleles were associated with anovulatory PCOS in two independent populations and were more frequent among women with Polycystic Ovaries (PCO) with symptoms than those without symptoms⁷⁵. In addition, it was shown that the fasting serum insulin levels were significantly higher in families with evidence of linkage⁷⁵. This evidence stands for the assumption that VNTR polymorphisms affect the presence of hyperinsulinemia and insulin resistance in some PCOS phenotypes. It was also reported that class III alleles were transmitted significantly more common from fathers than from mothers to affected daughter suggesting a "parent of origin" effect^{75, 76}. In support of this evidence, it was demonstrated that class III alleles and paternal class III allele transmissions were significantly related to increased number of PCOS features and to reduced insulin sensitivity among women with PCOS⁷⁷. In other studies, however, it was not found any evidence for the linkage of INS and PCOS and for the association of the class III allele and of hyperandrogenemia^{78,79}. But there was a difference in these studies. The ultrasonographic criteria were more commonly used than the NIHCD criteria. These conflicting results may be explained by the variant selection criteria, the different ethnic and geographic distribution of studied patients, the selection bias and the small size of the samples.

The insulin receptor is a heterotetrameric glycoprotein comprised of two α and two β -subunits and is encoded by the insulin receptor gene (INSR) located at the chromosome 19⁸⁰. Many researchers have tried to explore whether the mutations of INSR could explain insulin resistance in PCOS. The first studies of sequencing the INSR, the tyrosine kinase domain of INSR and the mutations by molecular scanning of the entire coding region of INS did not reveal any mutations⁸¹⁻⁸³. More recently, a comprehensive study published by Urbanek et al.⁸⁴ demonstrated a linkage with PCOS 367 well-characterized families from Europe. Another study investigating a broad region of the chromosome 19p13.2 found strong evidence for association with D19S884, supporting thus the previous findings⁸⁴. In a recent study, Siegel et al.⁸⁵ examined an SNP at the tyrosine kinase domain of INSR and found an association in lean patients with PCOS. This SNP could be a susceptible variant for PCOS, or a result of linkage disequilibrium with another INSR polymorphism.

The activation of the insulin receptor after insulin binding requires the autophosphorylation of the β -subunit of the insulin receptor⁸⁶. The following tyrosine kinase activity produced after autophosphorylation phosphorylates insulin receptor substrates (IRS), such as IRS-1 and IRS-2⁸⁷. Then, IRS-1 and IRS-2 bind and activate downstream effectors, such as phosphoinositide 3-kinase, to promote the metabolic and mitogenic actions of insulin. When IRS-1 is dysfunctional, IRS-2 is

the main messenger for the intracellular transmission of the insulin signal but it demands higher insulin concentration for activation⁸⁸. Several polymorphisms of IRS1 and IRS2 genes (IRS1 and IRS2) have been implicated in insulin resistance. The Gly972Arg polymorphism for IRS-1 and Gly1057Asp for IRS-2 have been shown to increase susceptibility to type-2 diabetes mellitus^{89,90}. Initially, no difference could be found in the distribution of IRS-1 Gly972Arg and IRS-2 Gly1057Asp alleles in PCOS patients and controls^{45,91}; however, it was demonstrated that the Gly972Arg IRS-1 was more prevalent in insulin-resistant patients compared with the non-insulin resistant patients or controls. Many studies following failed to prove any strong relationship or confirmation of any possible correlation between polymorphisms of IRS-1 and IRS-2 and PCOS⁹¹⁻⁹³.

In a recent study Dilek et al.⁹⁴ reported a higher frequency of the Gly972Arg polymorphism for IRS-1 in women with PCOS. Furthermore, similar to previous studies^{91,93} they found that the Gly972Arg carriers were more obese, more insulin-resistant and had higher fasting insulin levels in comparison to other PCOS patients and controls⁹⁴. These investigators also studied the same PCOS patients for the potential differential effects of metformin therapy on the basis of IRS-1 genotype⁹⁵. Metformin administration resulted in lower LH, DHEAS, T, and fasting insulin levels and decreased insulin resistance and FAI in Gly972Arg-negative PCOS women more effectively and significantly when compared with the Gly972Arg-positive women⁹⁵. These findings could be considered a rough indicator of the relationship between the IRS-1 genotype and the insulin resistance phenotype of PCOS. Ertunc et al.⁹⁵ studied the hypothesis that a possible mechanism for the action of metformin may be augmentation of the tyrosine phosphorylation of the insulin receptor β -subunit and IRS proteins and the increase of the insulin-dependent and nondependent cellular glucose uptake through the family of glucose transporter proteins. They hypothesized that variant IRS-1 proteins could not transmit signals in order to increase the glucose uptake of muscle and adipose cells. This may be an explanation of the association of IRS-1 genotype with insulin resistance in some of PCOS patients. The IRS polymorphisms of these studies seem to be related mostly with insulin resistance rather than PCOS.

Calpain-10 is a cysteine protease that participates in insulin secretion and action⁹⁶, and genetic studies have shown that variation in the gene (CAPN10) encoding calpain-10 is associated with type-2 diabetes⁹⁷. There was an effort to determine whether variation in the CAPN10 is associated with quantitative traits related to the pathogenesis of PCOS and type-2 diabetes⁹⁸. It was found association between the 112/121 haplotype of this gene and higher insulin levels in African-American women and an increased risk of PCOS in both African-American and white women^{7,98}. Consecutive studies have had conflicting results about the relation of polymorphisms with PCOS⁹⁹⁻¹⁰¹.

Genes involved in energy homeostasis

During the last years it has been recognized that the adipose tissue is not only a connective tissue but is also one of the active endocrine organs which secretes a wide variety of products called adipocytokines¹⁰². As a large proportion of women with PCOS are overweight, obese and extremely obese some genes of the most popular adipocytokines have been investigated as candidate genes in the pathogenesis of PCOS. Sequencing the leptin gene in a small group of PCOS patients failed to detect any mutations of the coding exons¹⁰³. In this study, the leptin receptor gene was also sequenced and revealed previously identified amino acid variants in exons 2, 4, 12 and the pentanucleotide insertion in the 3'-untranslated region¹⁰³. However, the allele frequencies of these polymorphisms did not differ from those in the general population.

Recent studies have focused on two polymorphisms, T45G in exon 2 and G276T in intron 2. It was demonstrated that these polymorphisms associate with obesity, insulin resistance, and the risk of developing type-2 diabetes¹⁰⁴⁻¹⁰⁶. In a study investigating the relationship of PCOS with 15 genomic variants previously described to influence insulin resistance, obesity, and type-2 diabetes mellitus, there was no association between PCOS and these two common polymorphisms of the adiponectin gene¹⁰⁷. Panidis et al. investigated the possible association of the T45G adiponectin gene polymorphisms with PCOS¹⁰⁸. A significant difference was observed between the groups when genotypes GG and TG were assessed together¹⁰⁹. It was also showed that the carriers of the G allele had a tendency for lower serum adiponectin levels in PCOS group¹⁰⁹. More recently, the probability that the T45G and G276T polymorphisms of adiponectin gene could be associated with PCOS was disputed by two studies^{109,110}. Concluding, the adiponectin gene do not seem to play a causative role in the pathogenesis of PCOS, rather seem to reflect the severity of the syndrome, at least concerning the metabolic disturbances and to have a role in the phenotypic variability of PCOS.

Genes involved in chronic inflammation

Tumor necrosis factor (TNF)- α is a cytokine secreted by adipose tissue with an important role in insulin resistance¹¹¹. The polymorphisms in the TNF- α gene do not seem to have a key role in the etiology of PCOS. In one study the carriers of the mutation 308 A alleles showed increased androgen and 17-hydroxyprogesterone levels before and after GnRH stimulation¹¹². These data may be indicative of the hypothesis that TNF- α gene polymorphism might be a modifying factor for phenotypic features. Other genes involved in chronic inflammation, such as TNFR2 (type-2 TNF receptor) gene¹¹³, IL-6¹¹⁴, IL-6 signal transducer gp 130¹¹⁵, IL-6 receptor¹¹⁵ genes have also been investigated for association with PCOS, but without significant results.

Abnormalities in the coagulation and fibrinolytic pathways contribute to the development of cardiovascular disease in PCOS patients¹¹⁶. Elevated plasminogen

activator inhibitor-1 (PAI-1) levels are associated with increased cardiovascular risk and increased thrombogenic tendency. Women with PCOS also present an increased activity of PAI-1¹¹⁷. In order to investigate the role of the PAI-1 polymorphism in PCOS patients, the polymorphism 4G/5G which is associated with higher PAI-1 concentrations, was evaluated in Greek women with PCOS and it was found a higher frequency in PCOS women compared with controls¹¹⁸. It was also reported that PCOS women have higher levels of PAI-1 and that the presence of the 4G allele in the PAI-1 promoter region of the gene further increases the PAI-1 levels¹¹⁸.

In addition to the genes mentioned above, many different genes such as HSD3B2¹¹⁹, 17 α -hydroxysteroid dehydrogenases¹²⁰, dopamine receptor^{121,122}, IGF¹⁰⁷, aldosterone synthetase¹²³, paraoxonase¹²³, glycogen synthetase¹²⁴, resistin¹²⁵, apoprotein E¹²⁶ have been studied. Results were either controversial or without clear conclusions.

References

- Cooper HE, Spellacy WN, Prem KA, Cohen WD. Hereditary factors in the Stein-Leventhal syndrome. *Am J Obstet Gynecol.* 1968; 100: 371-387.
- Amato P and Simpson JL. The genetics of polycystic ovary syndrome. *Best Pract Res Clin Obstet Gynaecol.* 2005; 18: 707-718.
- Wilroy RS Jr, Givens JR, Wiser WL, Coleman SA, Andersen RN, Summitt RL. Hyperthecosis: an inheritable form of polycystic ovarian disease. *Birth Defects:Original Article Series.* 1975; 11: 81-85.
- Givens JR. Familial polycystic ovarian disease. *Endocrinol Metab Clin N Am.* 1988; 17: 771-783.
- Franks S, Gharani N, Waterworth D, et al. The genetic basis of polycystic ovary syndrome. *Hum Reprod.* 1997; 12:2641-2648.
- Jahanfar S, Eden JA. Genetic and non-genetic theories on the etiology of polycystic ovary syndrome. *Gynecol Endocrinol.* 1996; 10: 357-364.
- Unluturk U, Harmanci A, Kocaefe C, Yildiz BO. The genetic basis of the polycystic ovary syndrome: a literature review including discussion of PPAR- γ . *PPAR Res.* 2007;1-23.
- Franks S, Gilling-Smith C, Gharani N, McCarthy M. Pathogenesis of polycystic ovary syndrome: evidence for a genetically determined disorder of ovarian androgen production. *Hum Fertil.* 2000; 3: 77-79.
- Gharani N, Waterworth DM, Batty S, et al. Association of the steroid synthesis gene CYP11a with polycystic ovary syndrome and hyperandrogenism. *Hum Mol Gen.* 1997; 6: 397-402.
- Diamanti-Kandaraki E, BartzisMI, Bergiele AT, Tsianateli TC, Kouli CR. Microsatellite polymorphism (ttta)n at -528 base pairs of gene CYP11a influences hyperandrogenemia in patients with polycystic ovary syndrome. *Fertil Steril.* 2000; 4: 735-741.
- Wang Y, Wu XK, Cao YX, et al. Microsatellite polymorphism of (ttta)n in the promoter of CYP11a gene in Chinese women with polycystic ovary syndrome. *Zhonghua Yi Xue Za Zhi.* 2005; 85: 3396-3400.
- Gaasenbeek M, Powell BL, Sovio U, et al. Large-scale analysis of the relationship between CYP11A promoter variation, polycystic ovarian syndrome, and serum testosterone. *J Clin Endocrinol Metab.* 2004; 89: 2408-2413.
- Tan L, Zhu G. Relationship between the microsatellite polymorphism of CYP11a gene and the pathogenesis of hyperandrogenism of polycystic ovary syndrome in Chinese. *Chin J Med Gen* 2005; 22: 216-218.
- Escobar-Morreale H, Pazos F, Potau N, Garcia-Robles R, Sancho JM, Varela C. Ovarian suppression with triptorelin and adrenal stimulation with adrenocorticotropin in functional hyperandrogenism: role of adrenal and ovarian cytochrome P450c17 α . *Fertil Steril.* 1994; 62: 521-530.
- Azziz R, Bradley EL Jr, Potter HD, Boots LR. Adrenal androgen excess in women: lack of a role for 17-hydroxylase and 17,20-lyase dysregulation. *J Clin Endocrinol Metab.* 1995; 80: 400-405.
- Witchel SF, Aston CE. The role of heterozygosity for CYP21 in the polycystic ovary syndrome. *J Pediatr Endocrinol Metab.* 2000; 13 (Suppl 5): 1315-1317.
- Witchel SF, Lee PA, Suda-Hartman M, Hoffman EP. Hyperandrogenism and manifesting heterozygotes for 21-hydroxylase deficiency. *Biochem Mol Med.* 1997; 62: 151-158.
- Escobar-Morreale HF, San Millan JL, Smith RR, Sancho J, Witchel SF. The presence of the 21-hydroxylase deficiency carrier status in hirsute women: phenotype/genotype correlations. *Fertil Steril.* 1999; 72: 629-638.
- Glintborg D, Hermann AP, Brusgaard K, Hangaard J, Hagen C, Andersen M. Significantly higher adrenocorticotropin-stimulated cortisol and 17-hydroxyprogesterone levels in 337 consecutive, premenopausal, caucasian, hirsute patients compared with healthy controls. *J Clin Endocrinol Metab.* 2005; 90: 1347-1353.
- Picado-Leonard J, Miller WL. Cloning and sequence of the human gene for P450c17 (steroid 17 α -hydroxylase/17,20-lyase): similarity with the gene for P450c21. *DNA.* 1987; 6: 439-448.
- Escobar-Morreale HF, Serrano-Gotarredona J, García-Robles R, Sancho JM, Varela C. Lack of an ovarian function influence on the increased adrenal androgen secretion present in women with functional ovarian hyperandrogenism. *Fertil Steril.* 1997; 67: 654-662.
- Wickenheisser JK, Quinn PG, Nelson VL, Legro RS, Strauss JF III, McAllister LM. Differential activity of the cytochrome P450 17 α -hydroxylase and steroidogenic acute regulatory protein gene promoters in normal and polycystic ovary syndrome theca cells. *J Clin Endocrinol Metab.* 2000; 85: 2304-2311.
- Wickenheisser JK, Nelson-Degrave VL, Quinn PJ, McAllister JM. Increased cytochrome P450 17 α -hydroxylase promoter function in theca cells isolated from patients with polycystic ovary syndrome involves nuclear factor-1. *Mol Endocrinol.* 2004; 18: 588-605.
- Wickenheisser JK, Nelson-DeGrave VL, McAllister JM. Dysregulation of cytochrome P450 17 α -hydroxylase messenger ribonucleic acid stability in theca cells isolated from women with polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2005; 90: 1720-1727.
- Carey AH, Waterworth D, Patel K, et al. Polycystic ovaries and premature male pattern baldness are associated with one allele of the steroid metabolism gene CYP17. *Hum Mol Gen.* 1994; 3: 1873-1876.
- Gharani N, Waterworth DM, Williamson R, Franks S. 5' polymorphism of the CYP17 gene is not associated with serum testosterone levels in women with polycystic ovaries. *J Clin Endocrinol Metab.* 1996; 81: 4174.
- Techatrasak K, Conway GS, Rumsby G. Frequency of a polymorphism in the regulatory region of the 17 α -hydroxylase-17,20-lyase (CYP17) gene in hyperandrogenic states. *Clin Endocrinol.* 1997; 46: 131-134.
- Witchel SF, Lee PA, Suda-Hartman M, Smith R, Hoffman EP. 17 α -hydroxylase/17,20-lyase dysregulation is not caused by mutations in the coding regions of CYP17. *J Pediatr Adol Gynecol.* 1998; 11: 133-137.
- Kahsar-Miller M, Boots LR, Bartolucci A, Azziz R. Role of a CYP17 polymorphism in the regulation of circulating dehydroepiandrosterone sulfate levels in women with polycystic ovary syndrome. *Fertil Steril.* 2004; 82: 973-975.

30. Simpson ER, Mahendroo MS, Means GD, et al. Aromatase cytochrome P450, the enzyme responsible for estrogen biosynthesis. *Endocrine Rev.* 1994; 15: 342-355.
31. Chen S, Besman MJ, Sparkes RS, et al. Human aromatase: cDNA cloning, Southern blot analysis, and assignment of the gene to chromosome 15. *DNA.* 1988; 7: 27-38.
32. Harada N, Ogawa H, Shozu M, Yamada K. Genetic studies to characterize the origin of the mutation in placental aromatase deficiency. *Am J Hum Gen.* 1992; 51: 666-672.
33. Ito Y, Fisher CR, Conte FA, Grumbach MM, Simpson ER. Molecular basis of aromatase deficiency in an adult female with sexual infantilism and polycystic ovaries. *Proceedings of the National Academy of Sciences of the United States of America.* 1993; 90: 11673-11677.
34. Erickson GF, Hsueh AJW, Quigley ME, Rebar RW, Yen SS. Functional studies of aromatase activity in human granulosa cells from normal and polycystic ovaries. *J Clin Endocrinol Metab.* 1979; 49: 514-519.
35. Jakimiuk AJ, Weitsman SR, Brzechffa PR, Magoffin DA. Aromatase mRNA expression in individual follicles from polycystic ovaries. *MolHum Reprod.* 1998; 4: 1-8.
36. Petry CJ, Ong KK, Michelmore KF, et al. Association of aromatase (CYP 19) gene variation with features of hyperandrogenism in two populations of young women. *Hum Reprod.* 2005; 20: 1837-1843.
37. Petry CJ, Ong KK, Michelmore KF, et al. Associations between common variation in the aromatase gene promoter region and testosterone concentrations in two young female populations. *J Ster Biochem Mo Biol.* 2006; 98: 199-206.
38. Lubahn DB, Joseph DR, Sullivan PM, Willard HF, French FS, Wilson EM. Cloning of human androgen receptor complementary DNA and localization to the X chromosome. *Science.* 1988; 240: 327-330.
39. Carson-Jurica MA, Schrader WT, O'Malley BW. Steroid receptor family: structure and functions. *Endocrine Rev.* 1990; 11: 201-220.
40. Chamberlain NL, Driver ED, Miesfeld RL. The length and location of CAG trinucleotide repeats in the androgen receptor N-terminal domain affect transactivation function. *Nucleic Acids Res.* 1994; 22: 3181-3186.
41. Giovannucci E, Stampfer MJ, Krithivas K, et al. The CAG repeat within the androgen receptor gene and its relationship to prostate cancer. *Proceedings of the National Academy of Sciences of the United States of America.* 1997; 94: 3320-3323.
42. Tut TG, Ghadessy FJ, Trifiro MA, Pinsky L, Yong LE. Long polyglutamine tracts in the androgen receptor are associated with reduced trans-activation, impaired sperm production, and male infertility. *J Clin Endocrinol Metab.* 1997; 82: 3777-3782.
43. Dowsing AT, Yong EL, Clark M, McLachlan RI, De Kretser DM, Trounson AO. Linkage between male infertility and trinucleotide repeat expansion in the androgen receptor gene. *Lancet.* 1999; 354: 640-643.
44. Mifsud A, Ramirez S, Yong EL. Androgen receptor gene CAG trinucleotide repeats in anovulatory infertility and polycystic ovaries. *J Clin Endocrinol Metab.* 2000; 85: 3484-3488.
45. Urbanek M, Legro RS, Driscoll DA, et al. Thirty-seven candidate genes for polycystic ovary syndrome: strongest evidence for linkage is with follistatin. *Proceedings of the National Academy of Sciences of the United States of America.* 1999; 96: 8573-8578.
46. Hickey T, Chandy A, Norman RJ. The androgen receptor CAG repeat polymorphism and X-chromosome inactivation in Australian Caucasian women with infertility related to polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2002; 87: 161-165.
47. Pugeat M, Crave JC, Tournaire J, Forest MG. Clinical utility of sex hormone-binding globulin measurement. *Horm Res.* 1996; 45: 148-155.
48. Selby C. Sex hormone binding globulin: origin, function and clinical significance. *An Clin Biochem.* 1990; 27: 532-541.
49. Berube D, Seralini GE, Gagne R, Hammond GL. Localization of the human sex hormone-binding globulin gene (SHBG) to the short arm of chromosome 17 (17p12→p13). *Cytogenet Cell Genet.* 1990; 54: 65-67.
50. Hogeveen KN, Talikka M, Hammond GL. Human sex hormone-binding globulin promoter activity is influenced by a (TAAAA)_n repeat element within an Alu sequence. *J Biol Chem.* 2001; 276: 36383-36390.
51. Xita N, Tsatsoulis A, Chatzikyriakidou A, Georgiou I. Association of the (TAAAA)_n repeat polymorphism in the sex hormone-binding globulin (SHBG) gene with polycystic ovary syndrome and relation to SHBG serum levels. *J Clin Endocrinol Metab.* 2003; 88: 5976-5980.
52. Cousin P, Calemard-Michel L, Lejeune H, et al. Influence of SHBG gene pentanucleotide TAAAA repeat and D327N polymorphism on serum sex hormone-binding globulin concentration in hirsute women. *J Clin Endocrinol Metab.* 2004; 89: 917-924.
53. Furui K, Suganuma N, Tsukahara SI, et al. Identification of two point mutations in the gene coding luteinising hormone (LH) β -subunit, associated with immunologically anomalous LH variants. *J Clin Endocrinol Metab.* 1994; 78: 107-113.
54. Okuda K, Yamada T, Imoto H, Komatsubara H, Sugimoto O. Antigenic alteration of an anomalous human luteinizing hormone caused by two chorionic gonadotropin type amino-acid substitutions. *Biochem Biophys Res Commun.* 1994; 200: 584-590.
55. Haavisto AM, Pettersson K, Bergendahl M, Virkamaki A, Huhtaniemi I. Occurrence and biological properties of a common genetic variant of luteinizing hormone. *J Clin Endocrinol Metab.* 1995; 80: 1257-1263.
56. Rajkhowa M, Talbot JA, Jones PW, et al. Prevalence of an immunological LH β -subunit variant in a UK population of healthy women and women with polycystic ovary syndrome. *Clin Endocrinol.* 1995; 43: 297-303.
57. Nilsson C, Pettersson K, Millar RP, Coerver KA, Matzuk MM, Huhtaniemi IT. Worldwide frequency of a common genetic variant of luteinizing hormone: an international collaborative research. *Fertil Steril.* 1997; 67: 998-1004.
58. Ramanujam LN, Liao WX, Roy AC, Loganath A, Goh HH, Ng SC. Association of molecular variants of luteinizing hormone with menstrual disorders. *Clin Endocrinol.* 1999; 51: 243-246.
59. Elter K, Erel CT, Cine N, Ozbek U, Hacıhanefioglu B, Ertungaalp E. Role of the mutations Trp8 → Arg and Ile15 → Thr of the human luteinizing hormone β -subunit in women with polycystic ovary syndrome. *Fertil Steril.* 1999; 71: 425-430.
60. Knight PG, Glistler C. Potential local regulatory functions of inhibins, activins and follistatin in the ovary. *Reprod.* 2001; 121: 503-512.
61. Guo Q, Kumar TR, Woodruff T, Hadsell LA, De-Mayo FJ, Matzuk MM. Overexpression of mouse follistatin causes reproductive defects in transgenic mice. *Mol Endocrinol.* 1998; 12: 96-106.
62. Legro RS, Spielman R, Urbanek M, Driscoll D, Strauss JF III, Dunaif A. Phenotype and genotype in polycystic ovary syndrome. *Recent Prog Horm Res.* 1998; 53: 217-256.
63. Urbanek M, Wu X, Vickery KR, et al. Allelic variants of the follistatin gene in polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2000; 85: 4455-4461.
64. Calvo RM, Villuendas G, Sancho J, San Millan JL, Escobar-Morreale HF. Role of the follistatin gene in women with polycystic ovary syndrome. *Fertil Steril.* 2001; 75: 1020-1023.
65. Junien C, van Heyningen V. Report of the committee on the genetic constitution of chromosome 11. *Cytogenet Cell Genet.* 1990; 55: 153-169.
66. Kennedy GC, German MS, Rutter WJ. The minisatellite in

- the diabetes susceptibility locus IDDM2 regulates insulin transcription. *Nat Genet.* 1995; 9: 293-298.
67. Paquette J, Giannoukakis N, Polychronakos C, Vafiadis P, Deal C. The INS 5' variable number of tandem repeats is associated with IGF2 expression in humans. *J Biol Chem.* 1998; 273: 14158-14164.
 68. Bell GI, Selby MJ, Rutter WJ. The highly polymorphic region near the human insulin gene is composed of simple tandemly repeating sequences. *Nature.* 1982; 295: 31-35.
 69. Weaver JU, Kopelman PG, Hitman GA. Central obesity and hyperinsulinaemia in women are associated with polymorphism in the 5' flanking region of the human insulin gene. *Eur J Clin Invest.* 1992; 22: 265-270.
 70. Ong KKL, Phillips DI, Fall C, et al. The insulin gene VNTR, type 2 diabetes and birth weight. *Nat Genet.* 1999; 21: 262-263.
 71. Dunaif A, Segal KR, Futterweit W, Dobrjansky A. Profound peripheral insulin resistance, independent of obesity, in polycystic ovary syndrome. *Diabetes.* 1989; 38: 1165-1174.
 72. Dunaif A, Segal KR, Shelley DR, Green G, Dobrjansky A, Licholai T. Evidence for distinctive and intrinsic defects in insulin action in polycystic ovary syndrome. *Diabetes.* 1992; 41: 1257-1266.
 73. Holte J, Bergh T, Berne C, Wide L, Lithell H. Restored insulin sensitivity but persistently increased early insulin secretion after weight loss in obese women with polycystic ovary syndrome. *J Clin Endocrinol Metab.* 1995; 80: 2586-2593.
 74. O'Meara NM, Blackman JD, Ehrmann DA, et al. Defects in β -cell function in functional ovarian hyperandrogenism. *J Clin Endocrinol Metab.* 1993; 76: 1241-1247.
 75. Waterworth DM, Bennett ST, Gharani N, et al. Linkage and association of insulin gene VNTR regulatory polymorphism with polycystic ovary syndrome. *Lancet.* 1997; 349: 986-990.
 76. Eaves IA, Bennett ST, Forster P, et al. Transmission ratio distortion at the INS-IGF2 VNTR. *Nat Genet* 1999; 22: 324-325.
 77. Michelmore K, Ong K, Mason S, et al. Clinical features in women with polycystic ovaries: relationships to insulin sensitivity, insulin gene VNTR and birth weight. *Clin Endocrinol.* 2001; 55: 439-446.
 78. Calvo RM, Telleria D, Sancho J, San Millan JL, Escobar-Morreale HF. Insulin gene variable number of tandem repeats regulatory polymorphism is not associated with hyperandrogenism in Spanish women. *Fertil Steril.* 2002; 77: 666-668.
 79. Vankova M, Vrbkova J, Hill M, Cinek O, Bendlova B. Association of insulin gene VNTR polymorphism with polycystic ovary syndrome. *An New York Acad Sci.* 2002; 967: 558-565.
 80. Goldfine ID. The insulin receptor: molecular biology and transmembrane signalling. *Endocrine Rev.* 1987; 8: 235-255.
 81. Sorbara LR, Tang Z, Cama A, et al. Absence of insulin receptor gene mutations in three insulin-resistant women with the polycystic ovary syndrome. *Metabolism.* 1994; 43: 1568-1574.
 82. Conway GS, Avey C, Rumsby G. The tyrosine kinase domain of the insulin receptor gene is normal in women with hyperinsulinaemia and polycystic ovary syndrome. *Hum Reprod.* 1994; 9: 1681-1683.
 83. Talbot JA, Bicknell EJ, Rajkhowa M, Krook A, O'Rahilly S, Clayton RN. Molecular scanning of the insulin receptor gene in women with polycystic ovarian syndrome. *J Clin Endocrinol Metab.* 1996; 81: 1979-1983.
 84. Urbanek M, Woodroffe A, Ewens KG, et al. Candidate gene region for polycystic ovary syndrome on chromosome 19p13.2. *J Clin Endocrinol Metab.* 2005; 90: 6623-6629.
 85. Siegel S, Futterweit W, Davies TF, et al. A C/T single nucleotide polymorphism at the tyrosine kinase domain of the insulin receptor gene is associated with polycystic ovary syndrome. *Fertil Steril.* 2002; 78: 1240-1243.
 86. Hubbard SR, Wei L, Ellis L, Hendrickson WA. Crystal structure of the tyrosine kinase domain of the human insulin receptor. *Nature.* 1994; 372: 746-754.
 87. Orio F, Palomba S, Colao A. Cardiovascular risk in women with polycystic ovary syndrome. *Fertil Steril.* 2006; 86(Suppl 1): S20-21.
 88. White MF. IRS proteins and the common path to diabetes. *Am J Physiol Endocrinol Metab.* 2002; 283: E413-E422.
 89. Burks DJ, White MF. IRS proteins and beta-cell function. *Diabetes.* 2001; 50 (Suppl1): S140-S145.
 90. Jellema A, Zeegers MPA, Feskens EJM, Dagnelie PC, Mensink RP. Gly972Arg variant in the insulin receptor substrate-1 gene and association with type 2 diabetes: a metaanalysis of 27 studies. *Diabetologia.* 2003; vol:990-995.
 91. El Mkaem SA, Lautier C, Macari F, et al. Role of allelic variants Gly972Arg of IRS-1 and Gly1057Asp of IRS-2 in moderate-to-severe insulin resistance of women with polycystic ovary syndrome. *Diabetes.* 2001; 50: 2164-2168.
 92. Ehrmann DA, Tang X, Yoshiuchi I, Cox NJ, Bell GI. Relationship of insulin receptor substrate-1 and -2 genotypes to phenotypic features of polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2002; 87: 4297-4300.
 93. Villuendas G, Botella-Carretero JI, Roldan B, Sancho J, Escobar-Morreale HF, SanMillan JL. Polymorphisms in the insulin receptor substrate-1 (IRS-1) gene and the insulin receptor substrate-2 (IRS-2) gene influence glucose homeostasis and body mass index in women with polycystic ovary syndrome and non-hyperandrogenic controls. *Hum Reprod.* 2005; 20: 3184-3191.
 94. Dilek S, Ertunc D, Tok EC, Erdal EM, Aktas A. Association of Gly972Arg variant of insulin receptor substrate-1 with metabolic features in women with polycystic ovary syndrome. *Fertil Steril.* 2005; 84: 407-412.
 95. Ertunc D, Tok EC, Aktas A, Erdal EM, Dilek S. The importance of IRS-1 Gly972Arg polymorphism in evaluating the response to metformin treatment in polycystic ovary syndrome. *Hum Reprod.* 2005; 20: 1207-1212.
 96. Sreenan SK, Zhou YP, Otani K, et al. Calpains play a role in insulin secretion and action. *Diabetes.* 2001; 50: 2013-2020.
 97. Horikawa Y, Oda N, Cox NJ, et al. Genetic variation in the gene encoding calpain-10 is associated with type 2 diabetes mellitus. *Nat Genet.* 2000; 26: 163-175.
 98. Ehrmann DA, Schwarz PEH, Hara M, et al. Relationship of calpain-10 genotype to phenotypic features of polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2002; 87: 1669-1673.
 99. Gonzalez A, Abril E, Roca A, et al. Comment: CAPN10 alleles are associated with polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2002; 87: 3971-3976.
 100. Gonzalez A, Abril E, Roca A, et al. Specific CAPN10 gene haplotypes influence the clinical profile of polycystic ovary patients. *J Clin Endocrinol Metab.* 2003; 88: 5529-5536.
 101. Haddad L, Evans JC, Gharani N, et al. Variation within the type 2 diabetes susceptibility gene calpain-10 and polycystic ovary syndrome. *J Clin Endocrinol Metab.* 2002; 87: 2606-2610.
 102. Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. *J Clin Endocrinol Metab.* 2004; 89: 2548-2556.
 103. Oksanen L, Tiitinen A, Kaprio J, Koistinen HA, Karonen SL, Kontula K. No evidence for mutations of the leptin or leptin receptor genes in women with polycystic ovary syndrome. *Mol Hum Reprod.* 2000; 6: 873-876.
 104. Hu FB, Doria A, Li T, et al. Genetic variation at the adiponectin locus and risk of type 2 diabetes in women. *Diabetes.* 2004; 53: 209-213.
 105. Stumvoll M, Tschrirer O, Fritsche A, et al. Association of the T-G polymorphism in adiponectin (Exon 2) with obesity and insulin sensitivity: interaction with family history of type 2 diabetes. *Diabetes.* 2002; 51: 37-41.
 106. Menzaghi C, Ercolino T, Di Paola R, et al. A haplotype at the adiponectin locus is associated with obesity and other features of the insulin resistance syndrome. *Diabetes.* 2002; 51: 2306-2312.
 107. San Millan JL, Corton M, Villuendas G, Sancho J, Peral B, Escobar-Morreale HF. Association of the polycystic ovary

- syndrome with genomic variants related to insulin resistance, type 2 diabetes mellitus, and obesity. *J Clin Endocrinol Metab.* 2004; 89: 2640-2646.
108. Panidis D, Kourtis A, Kukuvtis A, et al. Association of the T45G polymorphism in exon 2 of the adiponectin gene with polycystic ovary syndrome: role of Δ 4-androstenedione. *Hum Reprod.* 2004; 19: 1728-1733.
109. Xita N, Georgiou I, Chatzikyriakidou A, et al. Effect of adiponectin gene polymorphisms on circulating adiponectin and insulin resistance indexes in women with polycystic ovary syndrome. *Clin Chem.* 2005; 51: 416-423.
110. Escobar-Morreale HF, Villuendas G, Botella-Carretero JI, et al. Adiponectin and resistin in PCOS: a clinical, biochemical and molecular genetic study. *Hum Reprod.* 2006; 21: 22570-2265.
111. Hotamisligil GS, Peraldi P, Budavari A, Ellis R, White MF, Spiegelman BM. IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α - and obesity-induced insulin resistance. *Sci.* 1996; 271: 665-668.
112. Escobar-Morreale HF, Calvo RM, Sancho J, San Millan JL. TNF- α and hyperandrogenism: a clinical, biochemical, and molecular genetic study. *J Clin Endocrinol Metab.* 2001; 86: 3761-3767.
113. Peral B, San Millan JL, Castello R, Moghetti P, Escobar-Morreale HF. Comment: the methionine 196 arginine polymorphism in exon 6 of the TNF receptor 2 gene (TNFRSF1B) is associated with the polycystic ovary syndrome and hyperandrogenism. *J Clin Endocrinol Metab.* 2002; 87: 3977-3983.
114. Villuendas G, San Millan JL, Sancho J, Escobar-Morreale HF. The -597 G \rightarrow and -174 G \rightarrow C polymorphisms in the promoter of the IL-6 gene are associated with hyperandrogenism. *J Clin Endocrinol Metab.* 2002; 87: 1134-1141.
115. Escobar-Morreale HF, Calvo RM, Villuendas G, Sancho J, San Millan JL. Association of polymorphisms in the interleukin 6 receptor complex with obesity and hyperandrogenism. *Obes Res.* 2003; 11: 987-996.
116. Yildiz BO, Haznedaroglu IC, Kirazli S, Bayraktar M. Global fibrinolytic capacity is decreased in polycystic ovary syndrome, suggesting a prothrombotic state. *J Clin Endocrinol Metab.* 2002; 87: 3871-3875.
117. Legro RS. Polycystic ovary syndrome and cardiovascular disease: a premature association? *Endocr Rev.* 2003; 24: 302-312.
118. Diamanti-Kandarakis E, Palioniko G, Alexandraki K, Bergiele A, Koutsouba T, Bartzis M. The prevalence of 4G5G polymorphism of plasminogen activator inhibitor-1 (PAI-1) gene in polycystic ovarian syndrome and its association with plasma PAI-1 levels. *Eur J Endocrinol.* 2004; 150: 793-798.
119. Nayak S, Lee PA, Witchel SF. Variants of the type II β -hydroxysteroid dehydrogenase gene in children with premature pubic hair and hyperandrogenic adolescents. *Mol Genet Metab.* 1998; 64: 184-192.
120. Moghrabi N, Hughes IA, Dunaif A, Andersson S. Deleterious missense mutations and silent polymorphism in the human 17β -hydroxysteroid dehydrogenase 3 gene (HSD17B3). *J Clin Endocrinol Metab.* 1998; 83: 2855-2860.
121. Legro RS, Muhleman DR, Comings DE, Lobo RA, Kovacs BW. A dopamine 3 receptor genotype is associated with hyperandrogenic chronic anovulation and resistant to ovulation induction with clomiphene citrate in female Hispanics. *Fertil Steril.* 1995; 63: 779-784.
122. Kahsar-Miller M, Boots LR, Azziz R. Dopamine D3 receptor polymorphism is not associated with the polycystic ovary syndrome. *Fertil Steril.* 1999; 71: 436-438.
123. Zhao SP, Tang XM, Shao DH, Dai HY, Dai SZ. Association study between a polymorphism of aldosterone synthetase gene and the pathogenesis of polycystic ovary syndrome. *Zhonghua Fu Chan Ke Za Zhi.* 2003; 38: 94-97.
124. Rajkhowa M, Talbot JA, Jones PW, Clayton RN. Polymorphism of glycogen synthetase gene in polycystic ovary syndrome. *Clin Endocrinol.* 1996; 44: 85-90.
125. Urbanek M, Du Y, Silander K, et al. Variation in resistin gene promoter not associated with polycystic ovary syndrome. *Diabetes.* 2003; 52: 214-217.
126. Heinonen S, Korhonen S, Hippelainen M, Hiltunen M, Manermaa A, Saarikoski S. Apolipoprotein E alleles in women with polycystic ovary syndrome. *Fertil Steril.* 2001; 75: 878-880.